

# Pharmacy IV Admixture QA Procedures

A guide to monitoring of work area environmental conditions, validation of aseptic technique, and sterility and pyrogen testing

**Note: Recommendations are based on U.S. Pharmacopeia General Chapter <797>, Pharmaceutical Compounding - Sterile Preparations, effective 1/1/04**

## Environmental Monitoring

Startup Protocol - Perform tests daily for not less than 10 days.

### Air Sampling

1. Carefully remove EnviroTest paddles from their protective tubes just before using. Place one (1) EnviroTest in each work area of each Laminar Air Flow Workbench (LAFW) or barrier isolator to be monitored. Position each EnviroTest at least 6 inches inside the work area. EnviroTests should be centered in work areas, with the agar surface perpendicular to the normal unimpeded air flow.

If the LAFW or isolator is located in a "clean room", place an additional (1) EnviroTest outside of the LAFW. A location inside the clean room but near the entrance is preferred.

2. Expose EnviroTest agar to airflow for approximately one (1) hour. If the agar appears excessively dry after one hour, decrease exposure time to one half (2) hour.
3. Replace paddles into protective tubes. Place completed gummed label on cap of EnviroTests.
4. Incubate at 30-35°C for a minimum of 48 hours. Record results in log. (0 CFU's is normal)

### Surface Testing

5. Remove one (1) EnviroTest paddle from protective tube. Gently press surface of agar against selected areas in the LAFW or isolator, at least 6 inches inside the work area. Note: Do not slide agar across work surface.
6. Replace paddle into protective tube. Place completed gummed label on cap of EnviroTest.
7. Incubate at 30-35°C for a minimum of 48 hours. Record results in log. (0 CFU's is normal)

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Maintenance Protocol - Perform tests at least monthly in areas used to prepare low and medium-risk compounds and at least once per week in areas used to compound high-risk preparations.

1. Follow steps 1 through 7 above. Reduce air monitoring to one (1) EnviroTest per LAFW or isolator.
2. Any EnviroTest that yields 2 or more Colony Forming Units (CFUs) should trigger a return to the Startup Protocol test rate. Monitoring at the Startup rate should continue until the EnviroTests yield < 2 CFUs.
3. Check the HEPA filter, room air sources, cleaning solutions, equipment used in the hood, and other possible sources of the increased contamination. Immediately begin appropriate decontamination procedures.

### **Optional: Sanitizing Agents**

1. Check sanitizing agents such as sterile 70% isopropyl alcohol (IPA) for microbial contamination not less than once per week.
2. Using a sterile needle and syringe, inject a 1 to 20 mL aliquot of the sanitizing agent into an appropriately sized vial or bag of GroMed TSB media. The ratio of test solution to TSB media should not exceed 1 to 1.
3. Incubate at 20-25°C for 14 days. Record results.

### **Personnel & Process Validation using Media-Fill Testing**

Startup Protocol - Use to: (1) validate current pharmacists and technicians who manipulate sterile IV admixtures and (2) validate each newly hired staff member before they begin performing any manipulation requiring impeccable aseptic technique.

1. Choose a procedure that most closely simulates frequently used, complex, manipulations that are or will be performed by the pharmacy staff member. Directions for Use (DFU) that come with GroMed Personal Aseptic Technique Tests, PATT and PATT2, are suggestions and can be modified to fit a particular pharmacy's policies.
2. **Low-Risk:** PATT kits (#GM7020 & GM7030) exceed the challenge level required for a media-fill test procedure for low-risk level compounding.
3. **Medium-Risk:** PATT kits (#GM7020 & GM7030) exceed the challenge level required for a media-fill test procedure for medium-risk level compounding.
4. **Medium-Risk Increased Challenge:** Perform the steps suggested in the Directions for Use for the PATT or PATT2. Using a standard sterile pharmacy tubing set, use gravity to transfer the contents of the bag of TSB into either an empty vial or bag.

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5. **Medium-Risk Process Simulation:** Pharmacies that routinely use automated compounders can simulate common, complex process. Use bags (up to 500 mL) and vials of GroMed TSB media to substitute for both large and small volume liquid components. Typical medium-risk simulations with GroMed media include creating TPN and cardioplegia admixtures.
6. **High-Risk #1:** A high-risk level media-fill test that follows the USP <797> example can be accomplished by dissolving 3 grams of Soybean-Casein Digest Media, GroMed #GM3000, in 100 mL of non-sterile water. Or by dissolving 3 grams of a common, dry, non-bacterial static, non-sterile pharmacy component like NaCl or lactose in 100 mL of GroMed TSB liquid media.
7. If using a common powder, start by placing the 3 grams of non-sterile powder in a non-sterile beaker. Add 100 mL of GroMed TSB media by cutting off the outlet port on the bag or removing the rubber stopper on a vial. Stir to dissolve the powder.
8. Follow the remaining steps found in USP <797>.
9. **High-Risk #2:** Follow all the steps described in the PATT or PATT 2 directions for use.
10. Prepare 100 mL of nonsterile TSB as described in step #6 above. Draw 10 mL of the nonsterile TSB into a 20 mL syringe. Attach a 0.2µ syringe filter and sterile needle to the syringe. Inject 10 mL of sterilized TSB into the bag. Label and incubate as described in the PATT or PATT2 directions for use.

Maintenance Protocol - Perform not less than every twelve (12) months for low-risk level tests and every six (6) months for medium and high-risk level tests, or whenever unacceptable technique is observed.

1. Duplicate the procedure(s) used to initially validate the staff member. The number of repetitions of the basic manipulation may be reduced during revalidation if approved by the pharmacy manager. The procedure must be successfully completed without contamination. Any validation procedure that yields a positive result must be repeated until no positives are observed.
2. Positives should be analyzed to determine the probable break in aseptic technique that lead to the contamination. Conduct additional training and supervise as necessary to improve the individual's aseptic technique.
3. Record results in the log provided in each GroMed case. Optional - Enter results in the individual's personnel record.

## Admixture End Product Testing - Sterility

Startup and Maintenance Protocol - Test targeted admixtures according to USP <797>, Sterility Test<sup>®</sup>. Minimum test frequency is determined by batch size, multiple patient risk, and time/temperature exposure. The membrane filtration method is the method of choice where feasible. **All sterility tests should begin within 60 minutes of admixture preparation.**

1. Choose the QuickTest, QT Junior, or QTMicro that will test the complete admixture with the least amount of manipulation.
2. Pass admixture to be tested through the filter. For maximum sensitivity to the many potential sources of contamination, all of the LVP & SVP components should pass through the filter either during or within 60 minutes after completion of the admixing procedure being validated. Follow the Directions For Use (DFUs) enclosed in each case of testers.
3. Introduce GroMed media into filter chamber. Note for QuickTests: Admixtures containing fat emulsions may cause the TSB media to appear slightly cloudy. If necessary, use the extra GroMed media provided to rinse the filter of residual fat emulsion.
4. Complete and attach the gummed label to the filter housing.
5. Incubate the QuickTest, QTJunior, or QTMicro at 22+/-2.5<sup>o</sup>C for not less than 14 days. If the media becomes turbid before 14 days the test is <sup>Δ</sup>Positive<sup>®</sup>. Further observation is not necessary.
6. Positive tests should be investigated to locate the probable source of the admixture contamination. To help determine the source of accidental contamination, a positive tester may be sent to a microbiology laboratory to perform a species identification of the microorganism.
7. A "Negative" test may contain slow growing fungi. A blind secondary culture test for slow growing fungi can be performed using Potato Dextrose Agar (PDA) slants, GroMed #GM4000. Follow the directions supplied in each box of PDA slants.

## Admixture End Product Testing - Endotoxins

Startup Protocol - Use to ensure that each lot of nonsterile drug components in inventory do not exceed specified endotoxin limits.

1. Review the PyroTest Directions for Use (DFU). Verify that (1) the PyroTest is appropriate per the "Intended Use", (2) the drug(s) to be tested are compatible with gel-clot endotoxin testing, and (3) if the U.S. Pharmacopeia has assigned an endotoxin release limit.

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2. Using the drug's master work sheet and preparation work sheet, compound a small quantity of the drug to be tested. Compound the most commonly prescribed concentration.
3. Follow the appropriate PyroTest dilution procedure. Incubate the Assay and Positive Control Vials undisturbed for 60 minutes at 37°C.
4. Positive results in the Assay Vial indicates the test sample contains more than the maximum allowable endotoxin.

Maintenance Protocol - Use the maintenance protocol after the nonsterile drug components in inventory have been tested and shown to contain endotoxin levels below acceptable levels.

1. Perform endotoxin test upon receipt of each new lot of bulk, nonsterile drug components.
2. Perform endotoxin test on each batch of finished sterile drug products if the batch size, multiple patient risk, or time/temperature exposure criteria are exceeded according to USP <797>, Bacterial Endotoxin (Pyrogen) Testing@.
3. Use professional judgement regarding endotoxin testing when the preparation of a drug product is rare and/or the medical need is immediate.
4. Carefully follow the PyroTest DFUs. Always check the U.S. Pharmacopeia for updates to a drug's endotoxin release limit. Incubate the Assay and Positive Control Vials undisturbed for 60 minutes at 37°C.
5. Follow pharmacy's policy for quarantine and release from quarantine of covered drug products.

# Pharmacy IV Admixture QA Procedures

## Product Needs Analysis

### Environmental Monitoring

#### EnviroTests needed to support Startup Protocol

Number of hoods/isolators to monitor = \_\_\_\_\_  
Times 3 EnviroTests per hood (2 air + 1 surface) X 3 = \_\_\_\_\_  
Plus 1 EnviroTest per clean room +1 = \_\_\_\_\_  
Times #\_\_\_\_\_ of days of initial monitoring X Days = \_\_\_\_\_ # Initial EnviroTests needed

#### EnviroTests needed to support Maintenance Protocol

Number of hoods/isolators to monitor = \_\_\_\_\_  
Times 2 EnviroTests per hood (1 air + 1 surface) X 2 = \_\_\_\_\_  
Plus 1 EnviroTest per clean room +1 = \_\_\_\_\_  
Times #\_\_\_\_\_ of monitoring days per month X Days/month = \_\_\_\_\_ #EnviroTests needed / month

### Personnel & Process Validation

#### GroMed media needed to support Startup and Maintenance Protocol

Number of Pharmacists & Technicians to validate = \_\_\_\_\_  
Low Risk: #\_\_\_\_\_ PATT Kits per validation = \_\_\_\_\_ PATT Kits needed  
Medium/High Risk: #\_\_\_\_\_ PATT Kits per validation = \_\_\_\_\_ PATT Kits needed  
Medium/High Risk: #\_\_\_\_\_ containers of media per validation = \_\_\_\_\_ Vials & \_\_\_\_\_ Bags needed

### Admixture End Product Testing - Sterility

#### QuickTest, Junior, & Micro filter units needed to support Startup and Maintenance Protocol

Total number of targeted admixtures prepared per month = \_\_\_\_\_

### Admixture End Product Testing - Endotoxin

#### PyroTests needed to support Startup Protocol

Number of in-stock drug components to test = \_\_\_\_\_

#### PyroTests needed to support Maintenance Protocol

Number finished batches/doses of drugs to test = \_\_\_\_\_